

PROCEEDINGS OF THE TWENTY-EIGHTH ANNUAL KECK RESEARCH SYMPOSIUM IN GEOLOGY

April 2015
Union College, Schenectady, NY

Dr. Robert J. Varga, Editor
Director, Keck Geology Consortium
Pomona College

Dr. Holli Frey
Symposium Convener
Union College

Carol Morgan
Keck Geology Consortium Administrative Assistant

Christina Kelly
Symposium Proceedings Layout & Design
Office of Communication & Marketing
Scripps College

*Keck Geology Consortium
Geology Department, Pomona College
185 E. 6th St., Claremont, CA 91711
(909) 607-0651, keckgeology@pomona.edu, keckgeology.org*

ISSN# 1528-7491

The Consortium Colleges

The National Science Foundation

ExxonMobil Corporation

**KECK GEOLOGY CONSORTIUM
PROCEEDINGS OF THE TWENTY-EIGHTH ANNUAL KECK
RESEARCH SYMPOSIUM IN GEOLOGY
ISSN# 1528-7491**

April 2015

Robert J. Varga
Editor and Keck Director
Pomona College

Keck Geology Consortium
Pomona College
185 E 6th St., Claremont, CA
91711

Christina Kelly
Proceedings Layout & Design
Scripps College

Keck Geology Consortium Member Institutions:

**Amherst College, Beloit College, Carleton College, Colgate University, The College of Wooster,
The Colorado College, Franklin & Marshall College, Macalester College, Mt Holyoke College,
Oberlin College, Pomona College, Smith College, Trinity University, Union College,
Washington & Lee University, Wesleyan University, Whitman College, Williams College**

2014-2015 PROJECTS

RESILIENCE OF ENDANGERED ACROPORA SP. CORALS IN BELIZE. WHY IS CORAL GARDENS REEF THRIVING?:

Faculty: LISA GREER, Washington & Lee University, HALARD LESCINSKY, Otterbein University, KARL WIRTH, Macalester College

Students: ZEBULON MARTIN, Otterbein University, JAMES BUSCH, Washington & Lee University, SHANNON DILLON, Colgate University, SARAH HOLMES, Beloit College, GABRIELA GARCIA, Oberlin College, SARAH BENDER, The College of Wooster, ERIN PEELING, Pennsylvania State University, GREGORY MAK, Trinity University, THOMAS HEROLD, The College of Wooster, ADELE IRWIN, Washington & Lee University, ILLIAN DECORTE, Macalester College

TECTONIC EVOLUTION OF THE CHUGACH-PRINCE WILLIAM TERRANE, SOUTH CENTRAL ALASKA:

Faculty: CAM DAVIDSON, Carleton College, JOHN GARVER Union College

Students: KAITLYN SUAREZ, Union College, WILLIAM GRIMM, Carleton College, RANIER LEMPERT, Amherst College, ELAINE YOUNG, Ohio Wesleyan University, FRANK MOLINEK, Carleton College, EILEEN ALEJOS, Union College

EXPLORING THE PROTEROZOIC BIG SKY OROGENY IN SW MONTANA: METASUPRACRUSTAL ROCKS OF THE RUBY RANGE

Faculty: TEKLA HARMS, Amherst College, JULIE BALDWIN, University of Montana

Students: BRIANNA BERG, University of Montana, AMAR MUKUNDA, Amherst College, REBECCA BLAND, Mt. Holyoke College, JACOB HUGHES, Western Kentucky University, LUIS RODRIGUEZ, Universidad de Puerto Rico-Mayaguez, MARIAH ARMENTA, University of Arizona, CLEMENTINE HAMELIN, Smith College

Funding Provided by:
Keck Geology Consortium Member Institutions
The National Science Foundation Grant NSF-REU 1358987
ExxonMobil Corporation

GEOMORPHOLOGIC AND PALEOENVIRONMENTAL CHANGE IN GLACIER NATIONAL PARK, MONTANA:

Faculty: KELLY MACGREGOR, Macalester College, AMY MYRBO, LabCore, University of Minnesota

Students: ERIC STEPHENS, Macalester College, KARLY CLIPPINGER, Beloit College, ASHLEIGH, COVARRUBIAS, California State University-San Bernardino, GRAYSON CARLILE, Whitman College, MADISON ANDRES, Colorado College, EMILY DIENER, Macalester College

ANTARCTIC PLIOCENE AND LOWER PLEISTOCENE (GELASIAN) PALEOCLIMATE RECONSTRUCTED FROM OCEAN DRILLING PROGRAM WEDDELL SEA CORES:

Faculty: SUZANNE O'CONNELL, Wesleyan University

Students: JAMES HALL, Wesleyan University, CASSANDRE STIRPE, Vassar College, HALI ENGLERT, Macalester College

HOLOCENE CLIMATIC CHANGE AND ACTIVE TECTONICS IN THE PERUVIAN ANDES: IMPACTS ON GLACIERS AND LAKES:

Faculty: DON RODBELL & DAVID GILLIKIN, Union College

Students: NICHOLAS WEIDHAAS, Union College, ALIA PAYNE, Macalester College, JULIE DANIELS, Northern Illinois University

GEOLOGICAL HAZARDS, CLIMATE CHANGE, AND HUMAN/ECOSYSTEMS RESILIENCE IN THE ISLANDS OF THE FOUR MOUNTAINS, ALASKA

Faculty: KIRSTEN NICOLAYSEN, Whitman College

Students: LYDIA LOOPESKO, Whitman College, ANNE FULTON, Pomona College, THOMAS BARTLETT, Colgate University

CALIBRATING NATURAL BASALTIC LAVA FLOWS WITH LARGE-SCALE LAVA EXPERIMENTS:

Faculty: JEFF KARSON, Syracuse University, RICK HAZLETT, Pomona College

Students: MARY BROMFIELD, Syracuse University, NICHOLAS BROWNE, Pomona College, NELL DAVIS, Williams College, KELSA WARNER, The University of the South, CHRISTOPHER PELLAND, Lafayette College, WILLA ROWEN, Oberlin College

FIRE AND CATASTROPHIC FLOODING, FOURMILE CATCHMENT, FRONT RANGE, COLORADO:

Faculty: DAVID DETHIER, Williams College, WILLIAM B. OUMET, University of Connecticut, WILLIAM KASTE, The College of William and Mary

Students: GREGORY HARRIS, University of Connecticut, EDWARD ABRAHAMS, The College of William & Mary, CHARLES KAUFMAN, Carleton College, VICTOR MAJOR, Williams College, RACHEL SAMUELS, Washington & Lee University, MANEH KOTIKIAN, Mt. Holyoke College

SOPHOMORE PROJECT: AQUATIC BIOGEOCHEMISTRY: TRACKING POLLUTION IN RIVER SYSTEMS

Faculty: ANOUK VERHEYDEN-GILLIKIN, Union College

Students: CELINA BRIEVA, Mt. Holyoke College, SARA GUTIERREZ, University of California-Berkeley, ALESIA HUNTER, Beloit College, ANNY KELLY SAINVIL, Smith College, LARENZ STOREY, Union College, ANGEL TATE, Oberlin College

Funding Provided by:
Keck Geology Consortium Member Institutions
The National Science Foundation Grant NSF-REU 1358987
ExxonMobil Corporation

Keck Geology Consortium: Projects 2014-2015
Short Contributions— Belize Reef Project

MULTI-LEVEL CHARACTERIZATION OF ACROPORID CORAL POPULATIONS AT CORAL GARDENS, BELIZE: A REFUGIA IDENTIFIED

LISA GREER, Washington & Lee University, HALARD LESCINSKY, Otterbein University, KARL WIRTH, Macalester College

ARE THREESpot DAMSELFISH HELPING OR HURTING THE POSSIBLE RESURGENCE OF ACROPORA CORALS?

ZEBULON MARTIN, Otterbein University
Research Advisor: Dr. Halard Lescinsky, Otterbein University

GEOEYE-1 IMAGERY CLASSIFICATION: AN ACCURATE METHOD FOR IDENTIFYING POPULATIONS OF *ACROPORA* SPP. CORALS PRIOR TO A FIELD STUDY

JAMES BUSCH, Washington & Lee University
Research Advisor: Lisa Greer, Washington & Lee University

MORPHOMETRIC AND TAPHONOMIC ANALYSIS OF *ACROPORA PROLIFERA* AT CORAL GARDENS, BELIZE

SHANNON DILLON, Colgate University
Research Advisor: Constance M. Soja, Colgate University

***ACROPORA CERVICORNIS* RUBBLE AND FOSSIL FRAMEWORK AT CORAL GARDENS, BELIZE: INVESTIGATING ENVIRONMENTAL CONDITIONS AND SAMPLING STRATEGIES USING STABLE ISOTOPE GEOCHEMISTRY**

SARAH HOLMES, Beloit College
Research Advisor: Carl Mendelson, Beloit College

QUANTIFYING THE MICRO- AND MACRO- BORING COMMUNITIES IN CORAL GARDENS, BELIZE

GABRIELA GARCIA, Oberlin College
Research Advisor: Dennis K. Hubbard, Oberlin College

GRAZER DYNAMICS ON AN ACROPORID PATCH REEF SYSTEM AND THEIR IMPLICATIONS FOR THE CARBONATE BUDGET AT CORAL GARDENS, BELIZE

SARAH K. BENDER, The College of Wooster
Research Advisor: Mark Wilson, The College of Wooster

***ACROPORA CERVICORNIS* CARBONATE PRODUCTION AT CORAL GARDENS, BELIZE: PREDICTING FUTURE REEF STABILITY**

ERIN PEELING, The Pennsylvania State University
Research Advisor: Tim Bralower, The Pennsylvania State University

Funding Provided by:
Keck Geology Consortium Member Institutions
The National Science Foundation Grant NSF-REU 1358987
ExxonMobil Corporation

USING SEDIMENTS AND SUBSTRATES TO INTERPRET REGIONAL HYDRODYNAMICS AND ECOLOGY OF CORAL GARDENS, BELIZE

GREGORY MAK, Trinity University

Research Advisor: Daniel J. Lehrmann, Trinity University

GROWTH PATTERNS OF ACROPORA CERVICORNIS AFFECTED BY CURRENTS AT CORAL GARDENS, BELIZE

THOMAS R. HEROLD, The College of Wooster

Research Advisor: Shelley Judge, The College of Wooster

INVESTIGATIONS OF RESILIENT ACROPORA COMMUNITIES IN BELIZE: RELATIVE AGING AND INTRASPECIFIC DIVERSITY CALCULATIONS OF SPECIES USING MICROSATELLITE MARKERS AND SOMATIC MUTATIONS

ADELE IRWIN, Washington and Lee University

Research Advisor: Lisa Greer, Washington & Lee University

RECORD OF ENVIRONMENTAL CHANGE IN CARRIBEAN CORAL REEFS: SCLEROCHRONOLOGY AND GEOCHEMISTRY OF *O. FAVEOLATA* AS A PALEOCLIMATE PROXY AT CORAL GARDENS AND ROCKY POINT, BELIZE.

ILIAN A. DECORTE, Macalester College

Research Advisor: Karl R. Wirth, Macalester College

Funding Provided by:
Keck Geology Consortium Member Institutions
The National Science Foundation Grant NSF-REU 1358987
ExxonMobil Corporation

GRAZER DYNAMICS ON AN ACROPORID PATCH REEF SYSTEM AND THEIR IMPLICATIONS FOR THE CARBONATE BUDGET AT CORAL GARDENS, BELIZE

SARAH K. BENDER, College of Wooster
Research Advisor: Mark Wilson

INTRODUCTION

A shift to macroalgae-dominated reefs from coral-dominated reefs has been documented via photographs and data from the 1980's to the present in the Caribbean. The new fast-paced algal growth on reefs can result in a decrease in live coral abundance via competition for space and blockage of space for new coral recruitment (Aronson et al., 2002). Reef herbivores can help control algae abundance on reefs, but die-offs of important species like urchins and parrotfish have been recently and historically documented (Aronson et al., 2002). Changing dynamics between corals and surrounding grazer populations due to overfishing of herbivorous fish and disease outbreaks amongst urchins like *Diadema sp.* have resulted in a decline in living coral in the Caribbean (including Belize) (Aronson et al., 2002).

Bioeroding grazers, such as the sea urchin, *Echinometra viridis*, and parrotfish (*Scaridae*), play a vital role in maintaining the overall health of coral reef systems. Brown-Saracino et al. (2007) found that sea urchins acted as a control on the macroalgae of heavily fished coral reefs in Belize, but could also be a threat to reefs when present in high densities, as they are exceptional bioeroders. Aronson et al. (2002) note that *E. viridis* populations are as abundant as 20-100 urchins/m² in Belize and Bellwood et al. (2004), McClanahan and Shaffir (1990), and Brown-Saracino et al. (2007) discovered that there were strong correlations between the percentage of live coral cover, macroalgae cover, and urchin densities on coral reefs.

In general, healthy populations of sea urchins and parrotfish help to keep coral cover high and

macroalgae cover low, sustaining healthy coral reefs. In this study, it was hypothesized that there would be strong positive correlations between urchin densities and live acroporid coral or macroalgae cover at Coral Gardens, Belize. Using photoanalysis and *in situ* ecological surveys, we compared grazer densities with the percentage of live coral at Coral Gardens and analyzed the correlations for significance.

We found that these relationships were not significant (or even positive) based on the data we collected at Coral Gardens. We concluded that our data were limited by our methodologies. The high urchin densities found at Coral Gardens, in comparison to similar studies, led me to suggest that not all of the macroalgae cover was accounted for through the two-dimensional photoanalysis. This method of data collection could not account for the internal complexity of the coral architecture, which had the capacity to 'hide' macroalgae from our census.

Finally, using standard bioerosion calculations for *E. viridis* and *Scaridae*, a gross carbonate bioerosion rate was determined for Coral Gardens. This rate, combined with the gross carbonate production rate for *A. cervicornis* at Coral Gardens (Peeling, this volume), establishes an overall carbonate budget for Coral Gardens, which can be used to determine whether the reef is in a state of net growth or net erosion.

METHODS

Using a one square meter quadrat, a team of student photographers documented live coral cover and urchin densities along five transects using SCUBA. Urchin surveys took into account urchin species, density,

test diameter (by three size categories) and any other observations. This was done for all five *Acropora cervicornis* patch transects.

Fish surveys were done along each transect following the belt transect method of the Atlantic Gulf Rapid Reef Assessment (AGRRA). This method consists of a diver swimming at a constant rate along a 30-meter long and two meter wide belt transect for about 6 minutes while counting fish ahead of the diver. A 1-meter T-bar was used to estimate width of survey area and to help estimate fish size.

To determine the percentage of live coral cover along transects at Coral Gardens, we used a computer program (MATLAB) to trace the live *A. cervicornis* in each quadrat photo. After completing each quadrat at each transect, we were able to determine which percentage of the photograph was live *A. cervicornis*. We used this measurement to get average live coral per transect and compared to data from 2012 and 2013. We also used these percentages to compare the health of the reef with other ecological factors like urchin densities and fish counts.

Coral Point Count (CPC), was used to determine benthic cover in the “other” parts of the reef that were not live *A. cervicornis*. 100 points were randomly distributed across each photoquadrat and what the point has landed on was identified. We mainly used CPC to determine macroalgae cover but we also compared live coral cover as determined in MATLAB and CPC and found that the numbers were very similar.

Finally, we used previously determined rates of bioerosion for sea urchins and parrotfish to calculate annual bioerosion rates per m² at Coral Gardens (Perry et al., 2012). The Atlantic and Gulf Rapid Reef Assessment uses a simple calculation to convert fish size to fish weight, which can be used to find the amount of bioerosion (in kg of CaCO₃ per year per m²) for each species of parrotfish in a reef area.

RESULTS

Photo analysis data were combined with urchin density data to compare important ecological variables in the summer of 2014. Figure 1 shows average coral

cover, average macroalgae cover, and average urchin count per m² per transect at Coral Gardens. At T1 and T2, urchins appear to decrease with increase in coral cover but T5 shows little to no urchin presence and high coral cover. This graph also shows that as urchins increase at any transect, macroalgae increases, and coral cover is lower. Further regression analysis (Table 1) confirmed that the apparent relationships were not statistically significant.

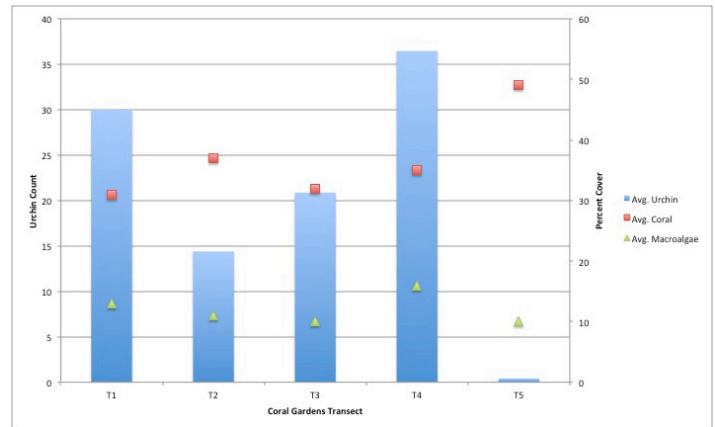


Figure 1. 2014 Coral Gardens average coral cover, average macroalgae cover, and average urchin count per m² per transect.

We found no significant correlation between urchin abundance and macroalgae cover at Coral Gardens (Fig. 2). A negative correlation exists between live coral cover and urchin abundance (Fig. 3). Further p-level analysis supported these findings (Table 1).

Transect	Urchin vs. Coral p-value	Hypothesis Rejected?	Urchin vs. Macroalgae p-value	Hypothesis Rejected?
T1	0.00312	No	0.33156	Yes
T2	0.00655	No	0.00272	No
T3	0.6772	Yes	0.01749	No
T4	0.2268	Yes	0.09021	Yes
T5	0.56713	Yes	0.73085	Yes

Table 1. Urchin, macroalgae cover, and coral cover correlation probability testing.

It is noteworthy that T1 and T2 had significant correlations between urchin density and coral cover. Transect 1 had the second-highest urchin density and the lowest percentage of live coral cover. Transect 2 had low urchin densities and the second highest percent coral cover.

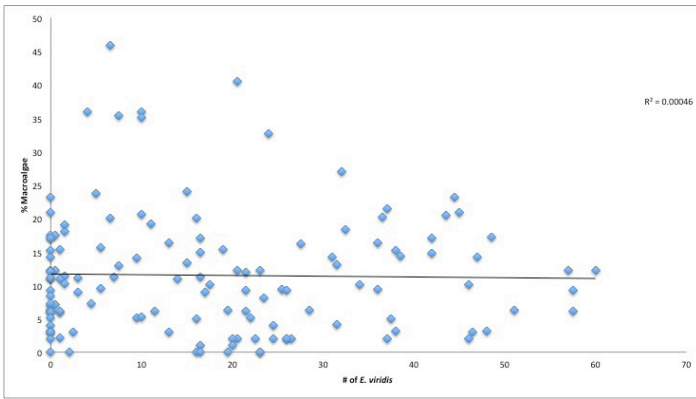


Figure 2. Urchin counts and macroalgae cover percentages at T1-T5. No correlation; hypothesis rejected.

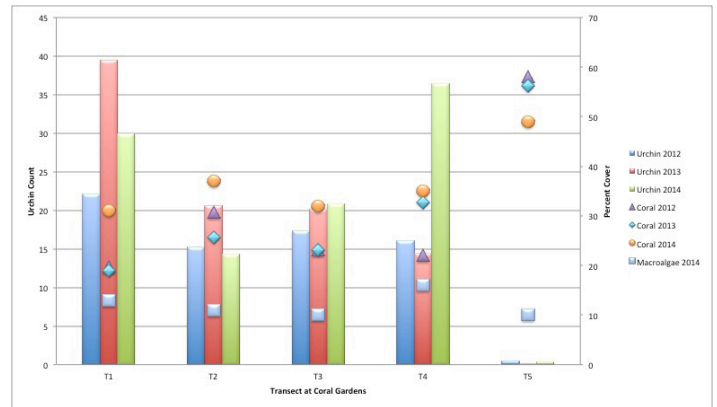


Figure 4. Average urchin densities per m^2 (Y_1) vs. average coral cover per m^2 (Y_2) across transects for all years and macroalgae cover for 2014.

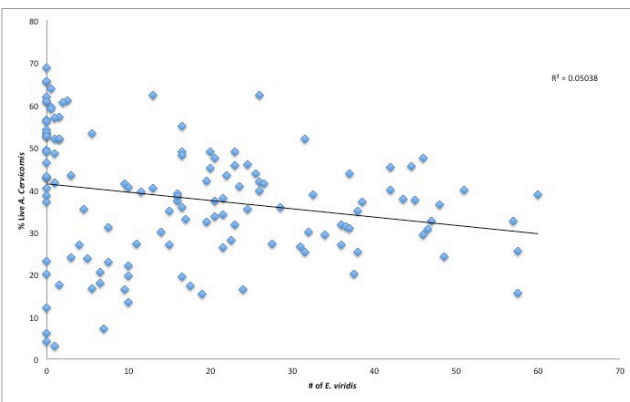


Figure 3. Urchin counts and live coral cover percentages at T1-T5. No correlation; hypothesis rejected.

An additional goal of this study was to determine the rate of bioerosion at each Coral Gardens transect using urchin and fish surveys and a previously determined rate of bioerosion for each species based on size (Perry et al., 2012). The total bioerosion rates for all fish and urchins at the five Coral Gardens transect are displayed in Table 2. Note the relatively high rates of bioerosion from parrotfish at Coral Gardens when compared to urchin bioerosion at the same transect.

To characterize the relationship of reef grazers and coral through time, we analyzed urchin and live coral cover data from 2012 and 2013 and compared it with data collected at Coral Gardens in 2014. Figure 4 shows how mean number of urchins, percent live coral, and macroalgae abundance per m^2 at each transect changed throughout the past three years. Generally, urchin densities increased from 2012 to 2013 and decreased into 2014. Transect 4 was an outlier, as it showed a slight decrease in urchin densities from 2012 and 2013 and a large increase in urchin densities into 2014. The coral cover does not reflect this increase, however, as we see it remains similar to the previous years. Another interesting trend in this graph is the increase in coral cover from 2013 into 2014 for Transect 1-3. Transect 5 is curious as it shows a decrease in coral cover in 2014.

	Urchin Count	Total Urchin Bioerosion (kg $CaCO_3/m^2/year$)	Parrotfish Count	Total Parrotfish Bioerosion (kg $CaCO_3/m^2/year$)
T1	960	0.412	7	0.178
T2	446	0.198	24	1.685
T3	396	0.287	7	1.186
T4	510	0.501	No data	No data
T5	15	0.006	19	3.791

Table 2. Bioerosion rates at Coral Gardens by transect. Values represent the total amount of calcium carbonate eroded by grazers in $kg CaCO_3/m^2/year$.

DISCUSSION

The data from this study indicates that, overall, there are no reliable relationships between live *Acropora cervicornis* and macroalgae in terms of cover and grazer populations at Coral Gardens. Many other studies in the Caribbean have found strong correlations between macroalgae, urchins, and coral cover (McClanahan, 1990). Because we found the opposite, it is important to look at the variations between these studies and ours.

Most modern studies of Caribbean reefs assess massive coral reefs (rather than branching coral in the case of our study) that are highly populated by macroalgae and less populated by urchins. At these massive coral-dominated reefs (such as *Montastrea*-dominated reefs), urchins are exposed at the top of the reef and must move around often to find food, making them easy targets for predators. Massive corals do not provide as much surface area for algae to flourish as branching corals, and urchins will leave if there is not enough food, leading to poor coral health and a transition to an algae-dominated reef.

At Coral Gardens, urchins like *E. viridis* have the opportunity to live within the large branching corals of the acroporids, as compared to other massive coral reefs. They are provided ample protection and food within the crevices of *A. cervicornis* and *A. palmata*. We therefore hypothesize that Coral Gardens is ecologically different from other reefs in the Caribbean and the relationship between bioeroders, live coral, and algae reflect these differences. Therefore we must be mindful of these ecological differences when comparing the relationships and correlations found at Coral Gardens to other studies on bioerosion or herbivory in the Caribbean.

Another conclusion that can be drawn from the lack of relationship between coral and macroalgae cover and sea urchins is that our data is limited by the fact that photo analysis does not account for three-dimensions of real-time data. It is highly likely that the photographs do not accurately represent the macroalgae present at Coral Gardens. We counted all of the urchins living inside the intertwined branches of the coral, but our photo analysis could not account for all of the macroalgae we observed living inside the

reef. Thus, it could be that there is enough macroalgae at Coral Gardens to keep the large urchin population fed, but it does not show in our data.

The unique inter-webbed living canopy of *A. cervicornis*–dominated reefs provides protection for urchins and sufficient surface area for macroalgae to exist on the dead branches (rubble) inside the reef. Because of the high acroporid coral cover at Coral Gardens, it may be concluded that the optimum density of *E. viridis* and *Scaridae*, as well as the optimum algal cover, is present here. The grazers provide room for new coral recruits of the fast-growing *A. cervicornis*, and the coral provides protection and food for the grazers.

Finally, no correlations were found between parrotfish and live coral or macroalgae cover. However, it is important to note the high bioerosion rates that scarids produce (in comparison to urchin bioerosion) at Coral Gardens. The total number of observed parrotfish at T1-T5 was 57. The total number of observed sea urchins at T1-T5 was over 2,000 (see Table 2). Yet, the rate of bioerosion from parrotfish was, on average, six times higher than that of urchins per transect. The presence of parrotfish at Coral Gardens, then, is highly influential on the carbonate budget of this reef and the presence of urchins has little effect. To support this claim, grazing and behavioral investigations of the parrotfish at Coral Gardens need to be conducted.

Coral Gardens is not a marine protected area (MPA), however, it lies between two other marine parks, Hol Chan Marine Reserve and Caye Caulker Marine Reserve, which may be responsible for the healthy grazer populations. Establishing protection for Coral Gardens could ensure an environment that fosters healthy parrotfish populations and continued high rates of acroporid coral growth and resilience.

CONCLUSIONS

- (1) According to our data, there is no significant correlation between sea urchins and live coral cover, or sea urchins and macroalgae cover at Coral Gardens. Our data is limited by the photo analysis method; I predict that more macroalgae is present at Coral Gardens than is recorded in the photos.

- (2) Coral Gardens displays a distinctive interlocking branching reef system. Fast-growing *A. cervicornis* provides protection and abundant food (macroalgae) for the grazer population, while grazers continuously “work” to open up space for new coral recruits. This is different than what we find elsewhere in Belize and the coral reef literature, thus traditional methods of measuring ecosystem health cannot be used.
- (3) Quantifying the fish densities at Coral Gardens and conducting behavioral investigations will be helpful in determining their effects on acroporid reef bioerosion.
- (4) Coral Gardens is in a state of net carbonate accretion based on the bioerosion and net growth calculations by Peeling (this volume). However, because we are assessing a different coral reef system and cannot properly quantify macroalgae using photoanalysis, the carbonate budget needs further analysis.
- (5) Establishing Coral Gardens as a Marine Protected Area (MPA) could help maintain current grazer-coral relationships (especially parrotfish populations), which may help play a part in sustaining its exceptional Acroporid health.

erosion: a new census-based carbonate budget methodology and applications to the reefs of Bonaire: *Coral Reefs*, v. 3, p. 853-868.

Perry, C. et al., 2013, Caribbean-wide decline in carbonate production threatens coral reef growth: *Nature Communications*, v. 4, p. 1-7.

REFERENCES

- Aronson, R., and Precht, W., 2002, White-band disease and the changing face of Caribbean coral reefs: *Hydrobiologia*, v. 460, p. 25-38.
- Bellwood, D.R., Hughes, T.P., Folke, C., and Nystro, M., 2004, Confronting the coral reef crisis: *Nature* v. 429, p. 827-833.
- Brown-Saracino, J., Peckol, P., Curran, H. A., and Robbart, M., 2007, Spatial variation in sea urchins, fish predators, and bioerosion rates on coral reefs of Belize: *Coral Reefs*, v. 26, p. 71-78.
- McClanahan, T. R. and Shafir, S.H., 1990, Causes and consequences of sea urchin abundance and diversity in Kenyan coral reef lagoons: *Oecologia*, v. 83, p. 362-370.
- Perry, C., et al., 2012, Estimating rates of biologically driven coral reef framework production and